

# Tox 21

## Developmental Toxicology

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The views expressed in this presentation are those of the presenter and do not necessarily reflect the views or policies of the US Environmental Protection Agency.

# Origins of Tox21

## “Toxicity Testing in the 21<sup>st</sup> Century”

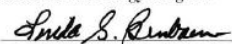


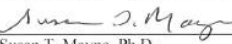
- Limitations in the current testing paradigm
  - Historical increase in:
    - Number of tests
    - Cost of testing
    - Use of laboratory animals
    - Time to develop and review data
  - Difficult to apply to risk assessment due to inability to fully address complex issues such as:
    - Life stage sensitivity
    - Mixtures and cumulative exposures
    - Varying exposure scenarios
    - Understanding of mechanism of toxicity and implications in assessing dose-response
    - Characterization of uncertainty



NRC, 2007

**NRC recommended a transformation in toxicity testing and risk assessment that focuses on toxicity pathways**

# Formation of U.S. Tox21 Federal Partnership - 2008

MEMORANDUM OF UNDERSTANDING	
ON	
High Throughput Screening, Toxicity Pathway Profiling, and Biological Interpretation of Findings	
<b>XI. APPROVAL</b>	
<b>National Toxicology Program</b>	
 Linda S. Birnbaum, Ph.D., DABT, ATS Director National Institute of Environmental Health Sciences National Institutes of Health	<u>5-11-15</u> Date
<b>National Center for Advancing Translational Sciences</b>	
 Christopher P. Austin, M.D. Director National Center for Advancing Translational Sciences National Institutes of Health	<u>5/20/2015</u> Date
<b>U.S. Environmental Protection Agency</b>	
 Fek G. Kadafi Acting Assistant Administrator Office of Research and Development U.S. Environmental Protection Agency	<u>6/16/15</u> Date
<b>U.S. Food and Drug Administration</b>	
 Susan T. Mayne, Ph.D. Director Center for Food Safety and Applied Nutrition U.S. Food and Drug Administration	<u>5/27/15</u> Date

- National Institute of Environmental Health Sciences (NIEHS)
- National Center for Advancing Translational Sciences (NCATS)
- U.S. Environmental Protection Agency (EPA)
- U.S. Food and Drug Administration (FDA)

## A Successful Interagency Collaboration:

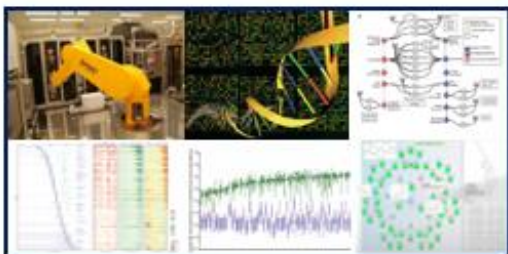
- Thousands of chemicals tested in 70 assays and over 50 relevant pathways (primarily HTP testing)
- Public release of millions of data points
- 200 peer reviewed articles in 56 journals
- Data now being used for regulatory decisions

# Tox21 Strategic and Operational Plan - 2018

## Areas of Focus

### Tox21 Collaboration

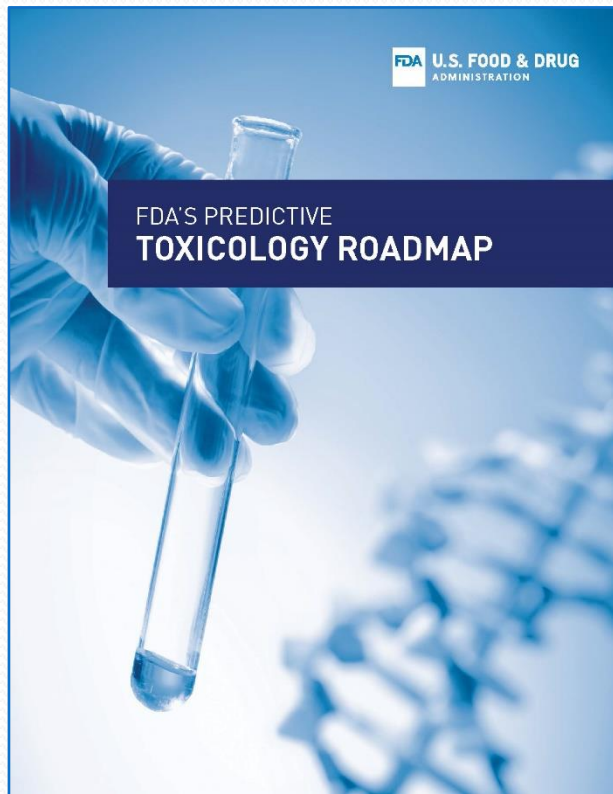
#### A Strategic Plan for Continued Leadership



- Developing and deploying alternative test systems that are predictive of human toxicity and dose response
- Addressing key technical limitations of current high throughput screening systems
- Consolidating chemical library management and developing more focused libraries
- Curating and characterizing legacy animal toxicity studies for continued comparison to high-throughput screening results
- Validating high-throughput assays, integrated assay batteries, computational models, 3-D organ-like model systems, and other emerging Tox21 approaches
- Refining and deploying high-throughput methods for characterizing pharmacokinetics to better predict the relationship between target tissue concentrations and external doses of chemicals.

# Agency-Specific Roadmaps Published

## FDA Predictive Toxicology Roadmap (Dec 2017)

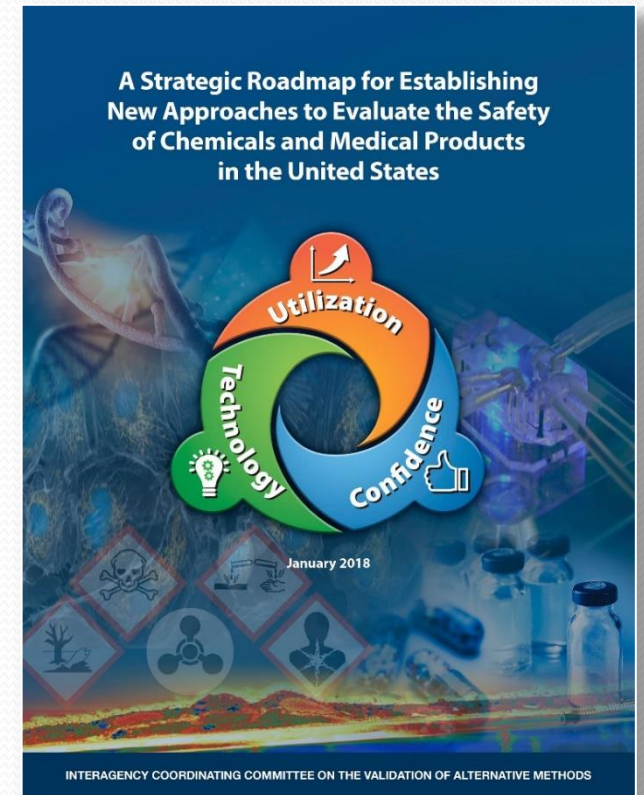


<https://blogs.fda.gov/fdavoices/index.php/2017/12/fda-launches-predictive-toxicology-roadmap-to-enable-advances-in-toxicity-testing/>

### Focus

- Reliable interpretation and application for product development and/or regulatory decisions
- Clear context of use
- Importance of multi-sector partnerships and collaborations to identify, develop, validate, and integrate assays into risk assessment

## ICCVAM Strategic Roadmap (Jan 2018)



<https://ntp.niehs.nih.gov/go/natl-strategy>

## Tox 21 Collaborative Projects

- Cell Line Selection for High-throughput Transcriptomics (Sipes, Harrill, Setzer)
- Acetylcholinesterase (AChE) Inhibitors Screening (Xia, Santillo)
- In Vitro Disposition of Tox21 Chemicals (DeVito, Friedman)
- High-Throughput (Ferguson, Harrill, Xia)
- Predictive Modeling of Developmental Toxicity with Human Pluripotent Stem Cells (Knudsen, Kleinstreuer, Lumen)
- Incorporating Genetic Susceptibility into Developmental Neurotoxicity Screening via Population Diversity (Harrill, Behl)
- Performance Based Validation of Tox21 Assays (Houck, Judson, Kleinstreuer)
- Retrofitting Existing Tox21 HTS Assays with Metabolic Capability (Xia, Witt, Simmons)

# An EPA Statutory Mandate for Chemical Testing

The 1976 Toxic Substances Control Act (TSCA) was amended by the **Frank R. Lautenberg Chemical Safety for the 21st Century Act** in 2016:

- new requirements and deadlines for actions related to the regulation of new and existing chemical substances.
- new subsection under Section 4 (*Testing of Chemical Substances and Mixtures*); particularly, Section 4 (h) entitled *Reduction of Testing on Vertebrates*

4(h)(2) - **Implementation of Alternative Testing Methods**—To promote the development and timely incorporation of new scientifically valid test methods and strategies that are not based on vertebrate animals, the Administrator **shall**—

4(h)(2)(A) - “not later than 2 years after the date of enactment....develop a strategic plan to **promote the development and implementation of alternative test methods and strategies** to reduce, refine, or replace vertebrate animal testing and provide information of equivalent or better scientific quality and relevance for assessing risks of injury to health or the environment of chemical substances or mixtures through, ...”

## Federal Register Notice, June 2015

### “Endocrine Disruptor Screening Program: Use of High Throughput Assays and Computational Tools”

<http://www.regulations.gov/#!documentDetail;D=EPA-HQ-OPPT-2015-0305-0001>

EDSP Tier 1 Battery of Assays	Model Alternative Development
Estrogen Receptor (ER) Binding	ER Model
Estrogen Receptor Transactivation (ERTA)	ER Model
Uterotrophic	ER Model
Androgen Receptor (AR) Binding	AR Model
Hershberger	AR Model
Aromatase	STR Model
Steroidogenesis (STR)	STR Model
Female Rat Pubertal	ER, STR & THY Models
Male Rat Pubertal	AR, STR & THY Models
Fish Short Term Reproduction	ER, AR & STR Models
Amphibian Metamorphosis	THY Model

<https://www.epa.gov/endocrine-disruption/use-high-throughput-assays-and-computational-tools-endocrine-disruptor>

<https://www.regulations.gov/docket?D=EPA-HQ-OPP-2017-0214>

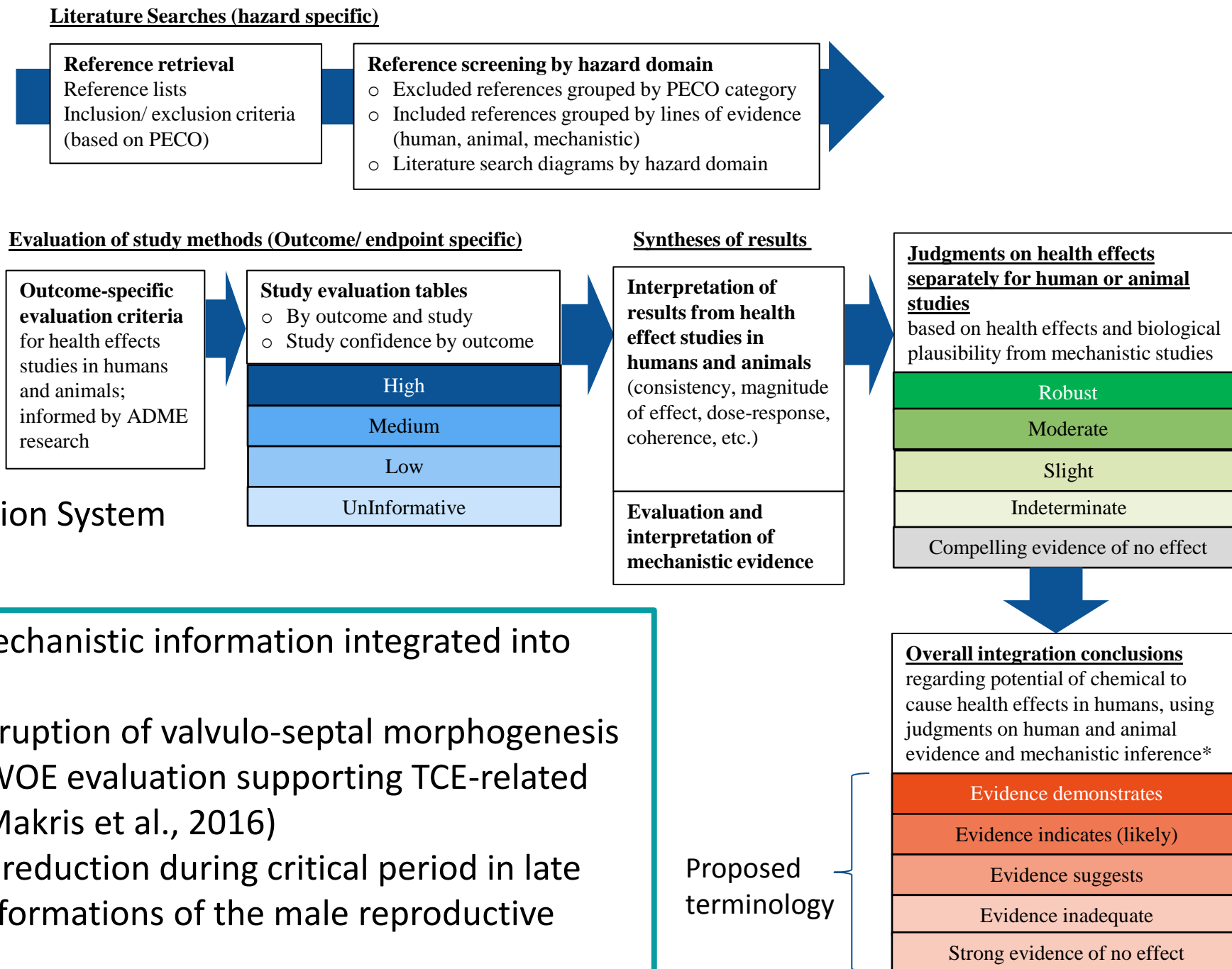


# Issues Typically Considered in the Evaluation and Interpretation of Regulatory Developmental Toxicity Data

- Bioavailability in the maternal animal (systemic exposure)
- Route(s) of exposure of the agent in vivo and any route-related differences in metabolism
- Ability of the agent or active metabolite to cross the placenta: fetal exposure
- Potential for the agent to cause death, altered growth structural abnormalities or functional effects in the offspring; effects of maternal toxicity or stress as a mitigating factor
- Life stage sensitivity: quantitative or qualitative effects
- Differences in toxicokinetic parameters (absorption, distribution, metabolism, storage, or excretion) in the fetus compared to adults
- Differences in toxicodynamic parameters in the fetus compared to adults: different targets or level of response
- Windows of susceptibility in the developing organism
- Dose-response (NOAELs/LOAELs/BMDs)

# EPA IRIS Systematic Review Process

IRIS = Integrated Risk Information System

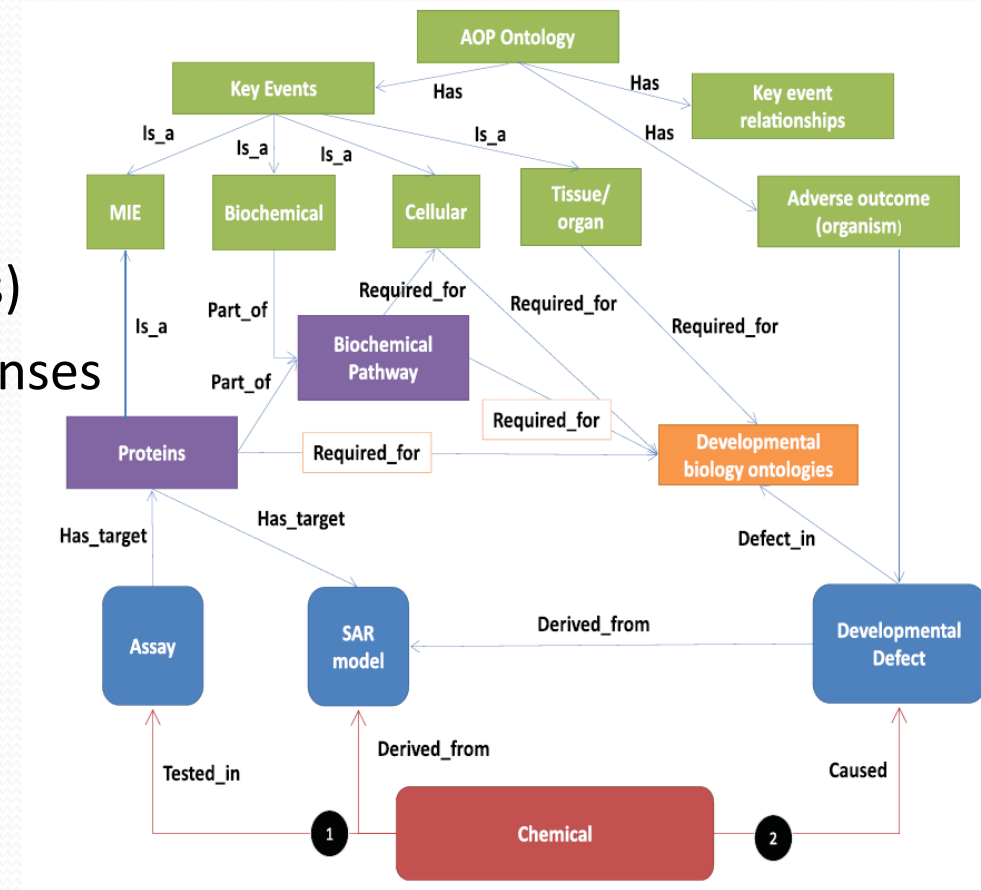


Examples of chemicals with mechanistic information integrated into devtox characterization:

- TCE: putative AOP for disruption of valvulo-septal morphogenesis was integrated into the WOE evaluation supporting TCE-related cardiac malformations (Makris et al., 2016)
- Phthalates: testosterone reduction during critical period in late gestation resulted in malformations of the male reproductive system

# Developmental Toxicity Ontology

- Ontology = a way to classify terms, how they relate to broader concepts and their interrelationships
- A developmental toxicity ontology can span multiple levels of organization and is based on:
  - Knowledge of developmental biology
  - Mode of action/ adverse outcome pathways
- Challenges
  - Role of potency (separating adaptive vs. adverse responses)
  - Maternal toxicity as a driver or confounder of in vivo responses
  - Importance of developmental stage susceptibility



DOI: 10.1002/bdr2.1189

ORIGINAL RESEARCH ARTICLE

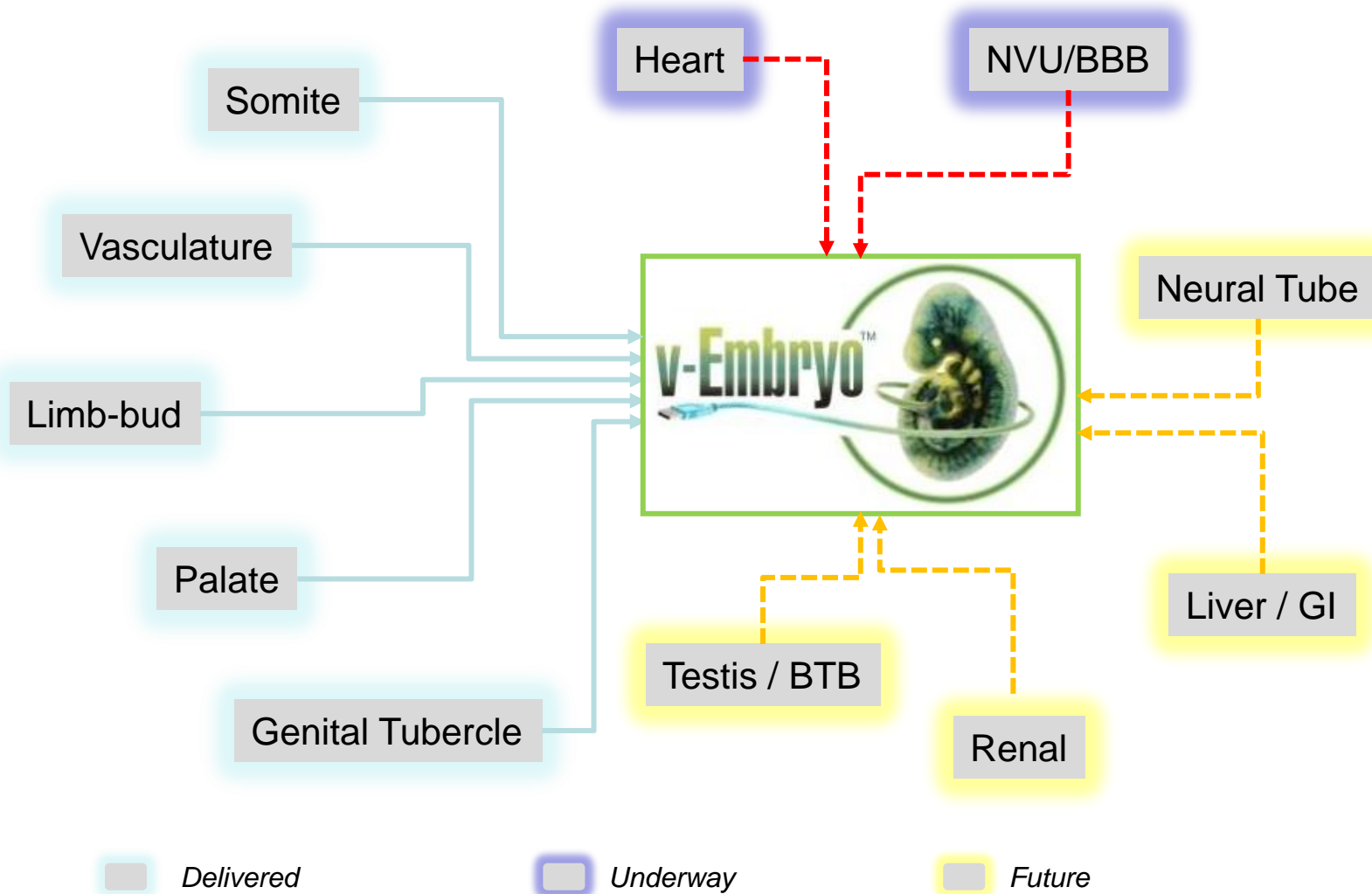


## Building a developmental toxicity ontology

Nancy Baker<sup>1</sup> | Alan Boobis<sup>2</sup> | Lyle Burgoon<sup>3</sup> | Edward Carney<sup>4\*</sup> |  
 Richard Currie<sup>5</sup> | Ellen Fritsche<sup>6</sup> | Thomas Knudsen<sup>7</sup> | Madeleine Laffont<sup>8</sup> |  
 Aldert H. Piersma<sup>9</sup> | Alan Poole<sup>8</sup> | Steffen Schneider<sup>10</sup> | George Daston<sup>11</sup>

Baker et al. 2018

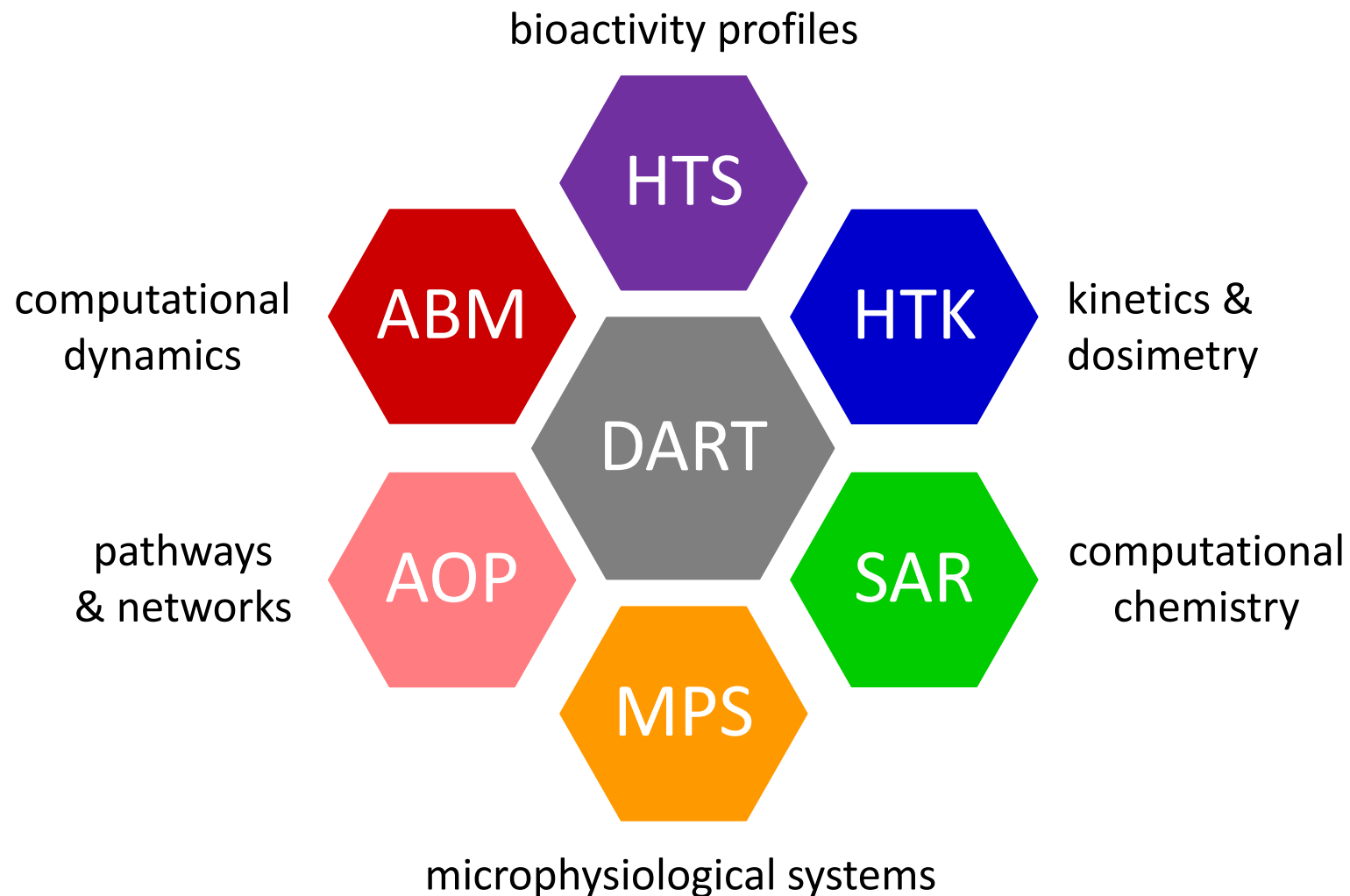
# Moving Toward a 'Virtual Embryo'



- Hester et al. (2011) PLoS Comp Bio; Dias et al (2014) Science
- Kleinstreuer et al. (2013) PLoS Comp Bio.
- Ahir et al. (MS in preparation)
- Hutson et al. (2017) Chem Res Toxicol.

- Leung et al. (2016) Reprod Toxicol.
- Zurlinden/Saili et al. (on-going)
- Hunter et al. (on-going)
- (Future research)

# IATA: Computational Synthesis and Integration



HTS – high throughput screen  
HTK – high throughput kinetics  
SAR – structure activity relationship  
MPS – microphysiological systems  
AOP – adverse outcome pathway  
ABM – agent based model

# Performance Check: In Vitro to In Vivo

- ToxCast\_STM anchored to 42 DevTox benchmark compounds aimed at assessing alternative models<sup>1</sup> and having information on pregnancy risk.
- Overall accuracy of 78.6% (0.65 sensitivity, 1.00 specificity, MCC = 0.647).
- Consistent with Palmer et al. (2013) pharma-trained model 77% accuracy (0.57 sensitivity, 1.00 specificity).

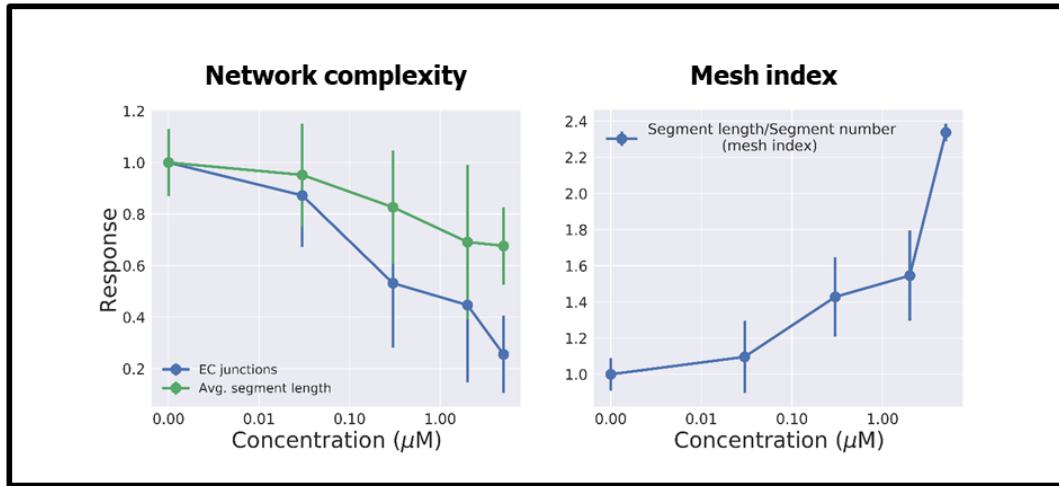
<sup>1</sup> Genschow et al. 2002; West et al. 2010; Daston et al. 2014; Augustine-Rauch et al. 2016; Wise et al. 2016

NCCT, manuscript in preparation

CASRN	Chemical	HTC <sup>1</sup> ( $\mu$ M)	CV <sup>2</sup> ( $\mu$ M)	TI <sup>3</sup> ( $\mu$ M)	Preg.class <sup>4</sup>	STM class <sup>5</sup>
302-79-4	all-trans-Retinoic acid	10	NA	0.003	X	TP
69-74-9	Cytarabine hydrochloride	1	0.083	0.054	D	TP
59-05-2	Methotrexate	1	0.062	0.059	X	TP
147-24-0	Diphenhydramine hydrochloride	100	3.76	0.588	B	TP
50-35-1	Thalidomide	100	NA	1.27	X	TP
51-21-8	5-Fluorouracil	1	1.45	2.02	D	TP
298-46-4	Carbamazepine	100	NA	2.29	C	TP
55-98-1	Busulfan	100	4.91	2.31	D	TP
13292-46-1	Rifampicin	10	NA	2.46	C	TP
19774-82-4	Amiodarone hydrochloride	10	NA	5.1	D	TP
75330-75-5	Lovastatin	20	NA	5.1	X	TP
3056-17-5	Stavudine	100	NA	32.5	C	TP
2392-39-4	Dexamethasone sodium phosphate	100	21.8	37.7	C	TP
53-86-1	Indomethacin	100	44.1	72.7	D	TP
127-07-1	Hydroxyurea	1000	237	74.9	D	TP
127-01-1	Valproic acid	1000	271	155	D	TP
4376-20-9	MEHP	500	NA	167	D	TP
57-41-0	5,5-Diphenylhydantoin	100	NA	NA	D	FN
51-52-5	6-Propyl-2-thiouracil	100	NA	NA	D	FN
10043-35-3	Boric acid	40.7	NA	NA	NTP	FN
4449-51-8	Cyclopamine	10	NA	NA	D	FN
6055-19-2	Cyclophosphamide monohydrate	20	NA*	NA	D	FN
56-53-1	Diethylstilbestrol	10	NA	NA	X	FN
107-21-1	Ethylene glycol	100000	NA	NA	NTP	FN
57-30-7	Phenobarbital sodium	100	NA*	NA	D	FN
81-81-2	Warfarin	100	NA	NA	X	FN
69-72-7	Salicylic acid	1000	1795	513	C	TN
103-90-2	Acetaminophen	100	NA*	NA	B	TN
79-06-1	Acrylamide	36	NA	NA	NTP	TN
50-78-2	Aspirin	100	NA*	NA	C	TN
80-05-7	Bisphenol A	100	39.4	NA	NTP	TN
94-26-8	Butylparaben	100	NA	NA	GRAS	TN
58-08-2	Caffeine	500	NA	NA	B	TN
464-49-3	D-Camphor	20	NA	NA	C	TN
131-11-3	Dimethyl phthalate	100	NA	NA	NTP	TN
59-30-3	Folic acid	100	NA	NA	A	TN
54-85-3	Isoniazid	8.8	NA*	NA	C	TN
57-55-6	1,2-Propylene glycol	1000000	246664	327552	NTP	TN
68-26-8	Retinol	10	NA	NA	A	TN
81-07-2	Saccharin	100	NA	NA	A	TN
134-03-2	Sodium L-ascorbate	20	NA*	NA	A	TN
599-79-1	Sulfasalazine	100	NA*	NA	B	TN
		True Positive Rate (sensitivity)		0.29	0.65	
		True Negative Rate (specificity)		0.94	1	
		Overall Accuracy		55.00%	78.60%	(MCC = 0.647)

# Performance Check: In Silico to In Vitro

## Computational prediction (cNVU)

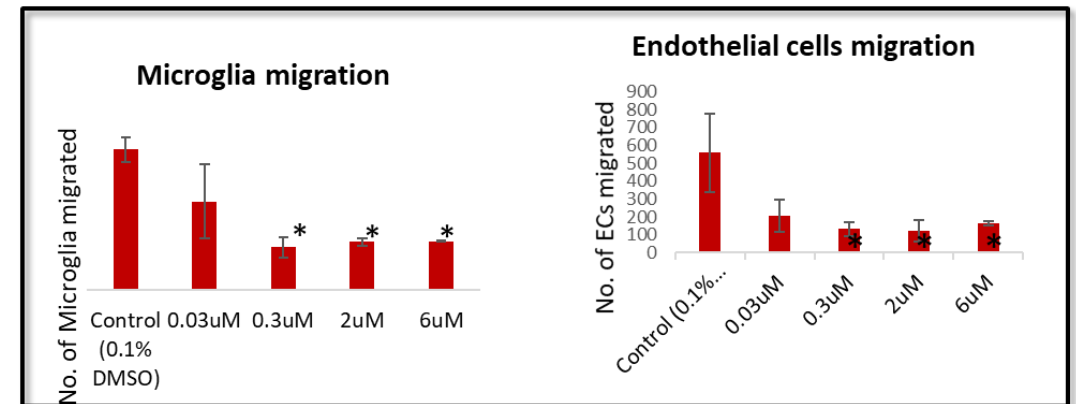
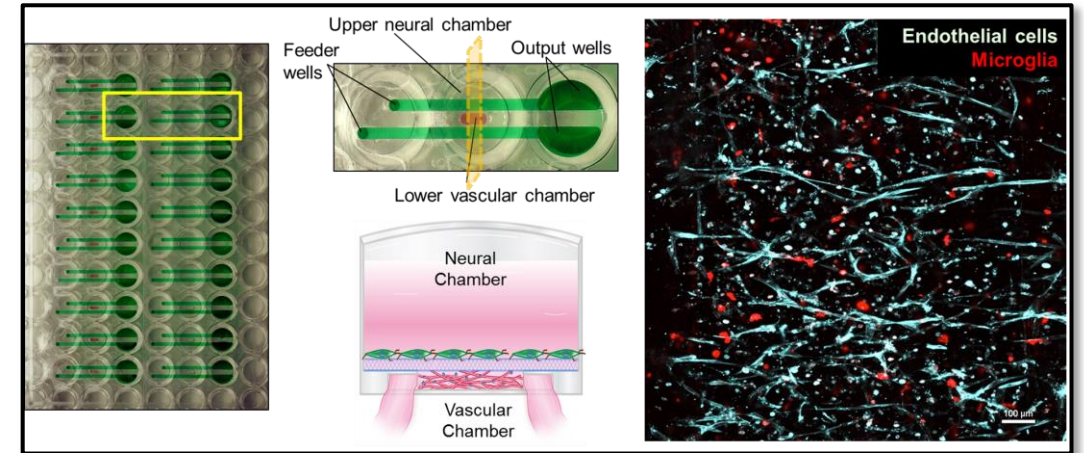


### Critical concentration:

- predicted *in silico*  $\sim 0.5 \mu\text{M}$
- observed *in vitro*  $\sim 0.3 \mu\text{M}$

Todd Zurlinden, Kate Saili - NCCT

## Biomimetic reconstruction (hNVU)



Murphy, W Daly, G Kaushick – U Wisconsin (HMAPS)

# Performance Check: Cross Assays

## Profiling the ToxCast library with pluripotent embryonic stem cell assays

Thomas B. Knudsen<sup>1</sup> Todd J. Zurlinden<sup>1</sup>, and E. Sidney Hunter<sup>2</sup>

U.S. EPA, Office of Research and Development <sup>1</sup>NCCT and <sup>2</sup>NHEERL

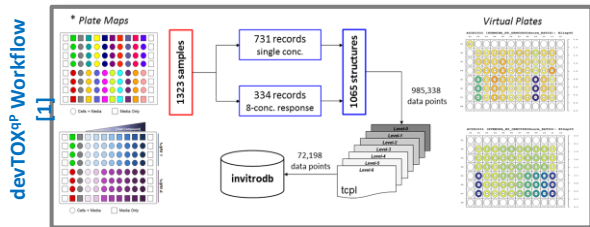
epa.gov | 919-541-9776  
1919-541-3490

### Introduction

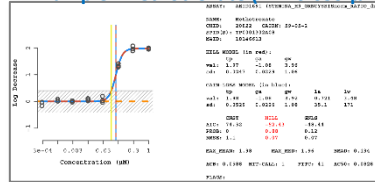
ToxCast chemicals were profiled for developmental toxicity potential in two embryonic stem cell assays and processed in the ToxCast data analysis pipeline (tcp1):

- [1] human pluripotent H9 stem cell-based (hESC) assay monitoring a metabolic biomarker [Palmer et al. 2013, BDRB];
- [2] mouse differentiating embryonic stem cell (mESC) adherent assay [Barrier et al. 2011, Reprod Tox]

### hESC (pluripotent) assay



#### Example: Methotrexate (TI = 0.059 μM)



- ↓ ornithine/cysteine in the day 3 secretome predicts μM threshold for teratogenicity (TI) [1];
- point of departure for cell viability equates to 11% reduction in cell number.

- TI was recorded for 181 chemicals (17% of 1065 tested); model performance used 42 benchmark compounds and ToxRefDB prenatal studies in rats and/or rabbits (dLEL ≤ 200 mg/kg/day) [manuscript in preparation].

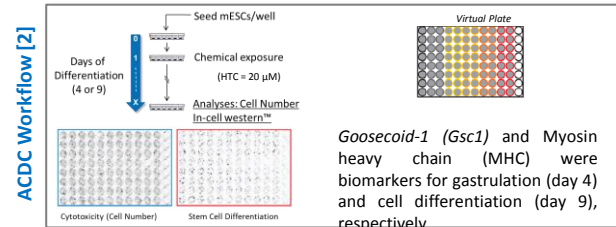
#### hESC model performance stringency filter applied to the in vivo anchor

	benchmark	none	low	medium	high
TP	17	85	60	35	19
FP	0	14	37	23	9
FN	9	217	127	51	11
TN	16	116	208	176	88
n	42	432	432	285	127
Sensitivity	0.654	0.281	0.321	0.407	0.633
Specificity	1.000	0.892	0.849	0.884	0.907
Accuracy	78.6%	46.5%	62.0%	74.0%	84.3%
MCC	0.647	0.190	0.202	0.332	0.554

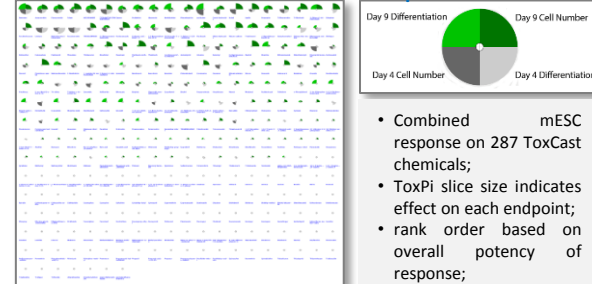
### Mining the ToxCast dataset to define assay sensitivity

To gain insight into the biological pathways and targets associated with the stem cell responses, machine-learning was used to mine correlations to 337 enzymatic and receptor signaling assays in the ToxCast NovaScreen dataset (NVS). Each NVS assay was enriched for an AC50 correlation against a hESC-positive or hESC-negative outcome, weighted by an assay-specific logistic regression model, processed through the Reactome HSA Pathway Browser (v3.5, database release 63), and independently enriched for significant pathway associations with the ClueGO plug-in to Cytoscape v3.4 (Bonferroni-corrected  $p \leq 0.05$ , minimum 3 genes for a pathway identifier).

### mESC (differentiation) assay



#### Effects of ToxCast chemicals on mESC endpoints



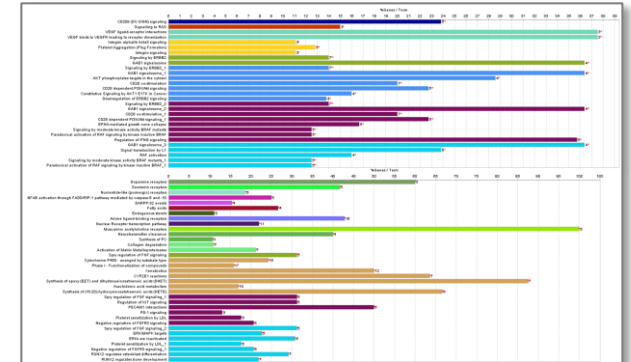
- Combined mESC response on 287 ToxCast chemicals;
- ToxPi slice size indicates effect on each endpoint;
- rank order based on overall potency of response;
- clustering reflects effect on the response pattern.

#### mESC model performance

- positive mESC response recorded on 95 chemicals (30.1% of 315 with ToxRefDB prenatal rat or rabbit studies);
- 221 dLEL-positives: *Gsc1* picked up 28% and MHC picked up 25% (overlap = 11% for *Gsc1* on day 4 + MHC on day 9).

Sensitive Domain (86 NVS targets)

Insensitive Domain (131 NVS targets)

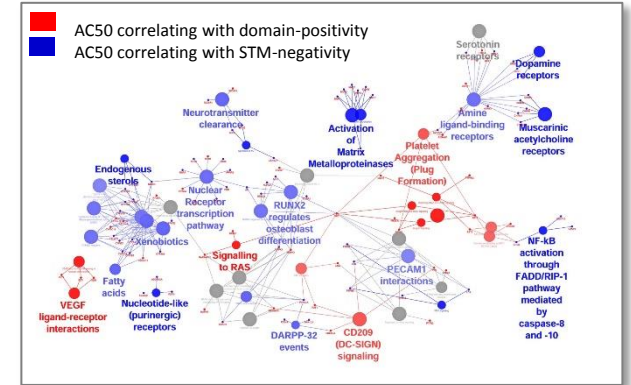


#### Domain (hESC): Network

- enriched pathway interactions mapped with the Cluepedia plug-in to Cytoscape.

- positive-response examples: inhibition of BRAF signaling, adrenocorticoids (GR, MR);

- negative-response examples: female hormone receptors (ESR1, PR), muscarinic receptors (M1-5), MMPs.



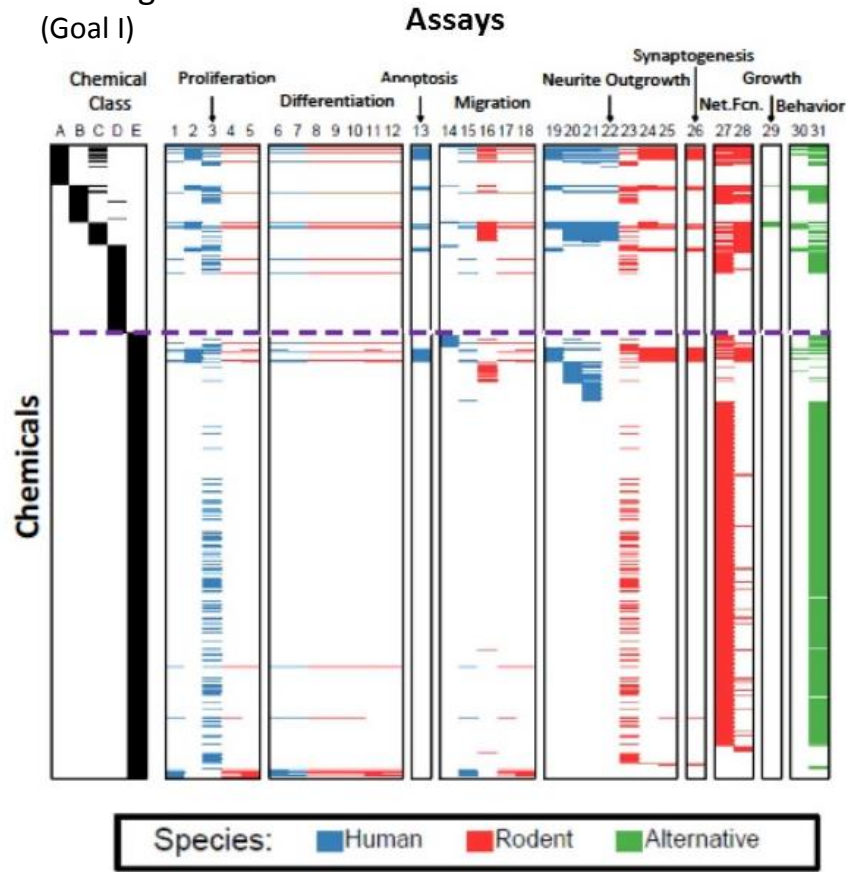
### Summary

- mESC examples: p53 signaling chemicals were classified for potential developmental toxicity using the hESC devTOX platform from Stemina Biomarker Discovery [1] or an adherent mESC assay [2].
- Performance against prenatal animal studies (ToxRefDB) improved from 62% to >84% accuracy as the level of confidence in the *in vivo* anchoring result (dLEL) increased.
- Characterizing the applicability domain at a pathway level sets the stage for new approach methodologies predicting developmental toxicity without vertebrate animal testing.

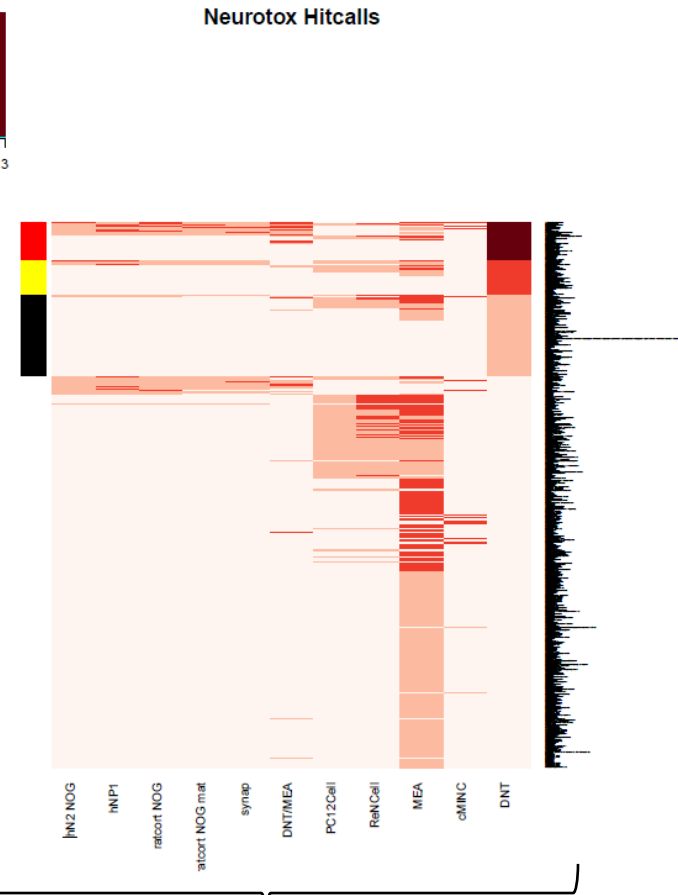
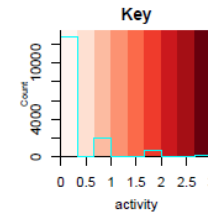


# EPA Developmental Neurotoxicity Research Plan

Compile Existing Data (Goal I)



GAPS  
Expand Chemical Universe (Goal II)



Data Translation (Goal III)

**Goal I: Compile existing data and identify gaps (focus on chemicals with in vivo data: Mundy et al. 2015)**

**Goal II: Expand the universe of compounds tested**

**Goal III: Data translation and accessibility**

**Goal IV: Provide a biological context for enhanced interpretation of DNT**

Neurotoxicology and Teratology 32 (2010) 26–38  
Contents lists available at ScienceDirect  
Neurotoxicology and Teratology  
journal homepage: www.elsevier.com/locate/neutera

Review article  
Expanding the test set: Chemicals with potential to disrupt mammalian brain development

William R. Mundy<sup>a,\*</sup>, Stephanie Padilla<sup>a</sup>, Joseph M. Breier<sup>a,1</sup>, Kevin M. Crofton<sup>b</sup>, Mary E. Gilbert<sup>a</sup>, David W. Herr<sup>a</sup>, Karl F. Jensen<sup>a</sup>, Nicholas M. Radio<sup>a,2</sup>, Kathleen C. Raffaele<sup>a</sup>, Kelly Schumacher<sup>a</sup>, Timothy J. Shafer<sup>a</sup>, John Cowden<sup>b</sup>

Source: Tim Shafer, EPA NHEERL

## Special Thanks

- Tom Knudsen (EPA/NCCT)
- Todd Zurlinden (EPA/NCCT)
- Katerine Saili (EPA/NCCT)
- Tim Shafer (EPA/NHEERL)
- Louis (Gino) Scarano (EPA/OPPT)
- Suzanne Fitzpatrick (FDA)

ToxCast/Tox21 data are located in the **CompTox Chemicals Dashboard**:  
<https://actor.epa.gov/dashboard/>